

The role of bioactive lipids in the development of soft tissue sarcomas in children – a review article

Rola bioaktywnych lipidów w rozwoju mięsaków u dzieci – przegląd piśmiennictwa

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SUMMARY

Soft tissue sarcoma accounts for almost 7% of all malignant tumours in children and adolescents. This heterogeneous group of mesenchymal and neuroectodermal tissue neoplasms can be characterized by common biological and clinics traits. Bioactive lipids are described as lipids, the levels of which – when

changed – lead to changes in cell functioning. The role of bioactive lipids in the biology of neoplastic cells is unquestionable. In this paper we present an overview of articles describing the impact of selected bioactive lipids on the development of sarcoma in children.

Key words: bioactive lipids, soft tissue sarcoma, children.

STRESZCZENIE

Mięsaki tkanek miękkich stanowią prawie 7% wszystkich złośliwych nowotworów rozpoznawanych u dzieci i młodzieży. Heterogeniczna grupa guzów rozwijająca się z tkanki mezenchymalnej i neuroektodermalnej charakteryzuje się zarówno podobnymi właściwościami biologicznymi, jak i cechami klinicznymi. Podkreśla się znaczenie lipidów bioaktywnych, ponieważ

każda zmiana ich stężenia powoduje zmianę w funkcjonowaniu komórki. Niezmiernie istotny jest wpływ bioaktywnych lipidów na funkcjonowanie komórek nowotworowych.

W prezentowanej pracy przedstawiono rolę wybranych bioaktywnych lipidów w rozwoju mięsaków tkanek miękkich u dzieci na tle światowego piśmiennictwa.

Słowa kluczowe: lipidy bioaktywne, mięsaki, dzieci.

INTRODUCTION

Soft tissue sarcoma (STS) accounts for almost 7% of all malignant tumours in children and adolescents. This heterogeneous group of mesenchymal and neuroectodermal tissue neoplasms can be characterized by common biological and clinics traits [1]. Rhabdomyosarcoma is the most common STS in children. It arises from progenitor cells of skeletal muscles and makes up to 50% of all described STS. Other malignant changes of soft tissue in children can be divided into two groups: RMS-like tumours (synovial sarcoma, primitive neuroectodermal tumour, extra-costal Ewing sarcoma, undifferentiated sarcoma) and non-RMS tumours [2].

Due to similarities in STS symptoms, location and histology it is often necessary to conduct genetic tests, which are crucial to establish the diagnosis. Good examples are alveolar RMS (ARMS), where in 70% of cases translocations t(2;13; p36; q14) can be found, leading to the presence of PAX3-FKHR and PAX7-FKHR fusion genes [3].

Treatment strategies have been evolving for 30 years and allow for good effects when combining chemotherapy and radiotherapy with surgical procedures. The mean five-year survival rate for all STS is from 50% to 80% including 70% in RMS, 50% in peripheral primitive neuroectodermal tumour

(PNET), and 80% in synovial sarcoma (SS). Upon establishing the diagnosis, distant metastases can be found in approximately 20% of patients. This finding decreases the effectiveness of treatment and makes for poorer prognosis. The mean five-year survival rate in those cases is 20–30%, but can be even lower for individuals with bone or bone marrow metastases [4].

Bioactive lipids are described as lipids, the levels of which – when changed – lead to changes in cell functioning. In the last two decades the most promising group of the abovementioned lipids comprise sphingolipids – present in every eukaryotic cell, and also in some prokaryotes [5]. The two substances of the large sphingolipid group discussed below are S1P and C1P. They are known to influence many cell functions, including those responsible for neoplastic growth. S1P stimulates cell proliferation and migration, leads to cytoskeletal actin rearrangement, and inhibits apoptosis. C1P increases inflammatory action by stimulating calcium-dependent mast-cell degranulation, and also increases cell survival by stimulating DNA synthesis [6, 7, 8]. Moreover, both of these bioactive lipids have been described as crucial to skeletal muscle development [9, 10].

This article aims to review the available literature dealing with the influence of bioactive lipids on the development and resistance to treatment of STS.

DISCUSSION AND REVIEW OF REFERENCES

A number of articles regarding the function of bioactive lipids in cell physiology during the development of various malignant tumours were reviewed. Special attention was paid to papers concerning STS in children.

Hokin and Hokin were the first to use the term "bioactive lipids" [11]. In the 1950s they observed and described a rapid turnover of inositol phospholipids in pancreatic slices that were stimulated with acetylcholine. Despite that fact, the role of diacereole (DAG) and inositol-1, 4, 5-triphosphate (Ins 1, 4, 5 P₃) in the regulation of protein kinase C (PKC) and calcium releasing was discovered later. DAG-PCK activation was reported, and it established the opinion that lipids can regulate cellular signals [5].

Sphingolipids are conserved in all eukaryotic cells and some prokaryotic cells (e.g. in the bacterial genus Sphingomonas). Loss of sphingolipid biosynthesis causes cell death in yeast, and CHO cell embryonic lethality in fly and mouse [12]. They are built of an amide-linked fatty acid chain and an amino-alcohol sphingosine base. Sphingolipids are irregularly distributed in the plasmatic membrane. Laterally, sphingolipids and cholesterol form membrane lipid microdomains. Vertically, sphingolipids and glycerophospholipids are unequally distributed between two leaflets of the plasmatic membrane [13, 14].

Ceramides are the central elements of sphingolipids metabolism and they can be generated by two major mechanisms. One of them is the *de novo* pathways of sphingolipids metabolism, and has a unique metabolic entry point, serine palmitoyl transferase (SPT), which forms the first sphingolipid of the novo pathway and as a unique exit point of this process, S1P lyase, which breaks down S1P into non-sphingolipid molecules. The second mechanism is known as the "sphingomyelin cycle", and it is responsible for the production of ceramide, which involves hydrolysis of the plasma's membrane sphingomyelin to ceramide and phosphorylcholine throughout the action of sphingomyelinase [15, 16]. In mammals S1P is described as acting as a receptor ligand, an intracellular signalling molecule, and the intermediate form in sphingolipid-to-glycerophospholipid conversion [12].

Up to now, five S1P receptors have been identified: from S1P₁ to S1P₅. The S1P₁ family receptors are G protein-coupled receptors (GPCRs) [17]. In mammals S1P₁, S1P₂ and S1P₃ are expressed in all tissues, whereas S1P₄ and S1P₅ are only restricted to some. S1P₄ is present in lymphoid tissue (thymus, spleen, lymph nodes), while S1P₅ is present in the brain and skin. When S1P is binding to its receptor, it activates a downstream signalling pathway leading to a variety of cellular responses, such as proliferation, regulation of cell migration and actin cytoskeletal rearrangement, and adherents junction assembly [18, 19]. There is also a specific role of S1P and S1P₁ in lymphocyte trafficking between the blood and lymphoid tissue [20, 21]. S1P is also known to be involved in the stimulation of chemotaxis cell survival and regulation of angiogenesis [22, 23].

C1P is reported to act as an intracellular signalling molecule. Up to now, no surface cell receptors for C1P have been

described despite many researches suggesting that C1P plays an important role in the immune system, and is an exogenous factor for cell survival and DNA synthesis, and that it increases intracellular Ca²⁺ concentrations [5, 7, 24].

Soft tissue sarcomas usually occur in infants 2 to 6 years old and teenagers 15 to 19 years old. These tumours can develop in various areas of the body: head, neck, upper and lower extremities, pelvic area, genitourinary system, and in the chest and lungs. Certain locations can be characteristic for particular histological types. Sarcomas are considered to be locally malignant with their expanding growth and frequent recurrences in the primary location. These tumours are also able to cause distant metastases by haematogenous spread into the lungs, bone marrow and nervous system [1, 2, 25].

Sarcomas, especially rhabdomyosarcoma, are more frequent in children with inborn defects of the central nervous and genitourinary systems, gastrointestinal tract and cardiac defects, as well as in patients suffering from neurofibromatosis, and in Li-Fraumeni Syndrome-affected families [26, 27, 28]. Clinical symptoms depend on the location of the neoplasm but not on histological type, and hence there are no symptoms characteristic for any particular sarcoma. Most of the conditions are caused by either mechanical compression of other anatomical structures or expansion of the tumour [1, 2, 25]. The diagnosis should be established based on a detailed medical history, and physical and radiological examinations, as well as laboratory and molecular tests. The cancer staging notation is usually established based on the size and radicality of the resection, and the presence of distant metastases and the tumour's response to chemotherapy [1, 3, 29].

Treatment strategies for sarcomas were developed by the Intergroup Rhabdomyosarcoma Study (IRS) and the International Society of Paediatric Oncology (SIOP - MMT), Cooperative Weichteilsarkom – studiengruppe – Gesellschaft für Pädiatrische Onkologie und Hämatologie, (CWS – GPOH), and the Italian Cooperative Study Group – Associazione Italiana Ematologia Oncologia Pediatrica (ICG – AIEOP). The aim of the treatment is to achieve the best possible control over the local and systemic development of the disease. The therapy combines surgical treatment with chemotherapy and radiotherapy [30, 31, 32, 33]. So far, there is only one report regarding the influence of S1P and C1P on the cells of STS in children. In that study Schneider *et al.* described the influence of S1P and C1P on rhabdomyosarcoma cell lines, including both ARMS (RH4, RH5, 4H18, RH28, RH30, RH41, CW9019) and ERMS (JR, SMS-CTR, RD, RH36) [34].

The influence of bioactive lipids was evaluated regarding RMS cell proliferation, chemotaxis, phosphorylation of intracellular protein pathways, and adhesion to fibronectin and bone marrow derived stroma cells, as well as neoplastic growth in immunodeficient mice. The lack of S1P and C1P influence on RMS cell proliferation was without any doubt very surprising when considering earlier reports pointing to their stimulating role in skeletal muscle cell development. Schneider *et al.* highlighted the considerable influence of various bioactive lipids on cell migration. Further research has allowed

them to establish that the activation of S1P₁ receptors on the surface of neoplastic cells is responsible for that fact. Moreover, inhibition of the S1P₁ receptor in these cells leads to the similar effect of increased cell mobility [35]. Another curious piece of information regards the discovery of increased RMS cell adhesion to fibronectin influenced by both S1P and C1P. The increased adherence to BM-derived stroma cells was observed under the influence of S1P only.

The authors considered the prometastatic effect of radiotherapy and chemotherapy as one of side effects, and they measured levels of S1P and C1P in the cells of tissues frequently affected by RMS metastases (bone marrow, liver, brain, lungs) before and after the treatment of immunodeficient mice implanted with RMS cells and subsequently treated with radiotherapy and chemotherapy. A significant increase of S1P and C1P was observed in bone marrow exposed to radiotherapy/chemotherapy. All those observations may point to S1P and C1P as one of the RMS cell mobility factors and for its ability to cause distant metastases as an effect of chemotherapy and radiotherapy. Curiously, the same group of researchers conducted similar experiment with other bioactive lipids, LPA and LPC, observing their influence on RMS cells with the same results as presented above. This not only suggests the identification of one factor promoting distant metastases, but also new possibilities of supporting therapy in treating cancer. Various publications regarding different points of the sphingolipid metabolism that could be influenced by exogenous agents confirm that.

Another group of researchers focused on the applicability of the S1P₁ receptor in the differentiation of neoplasms. It is known that this receptor can be found on the surface of endothelial cells. *Akiyama et al.* examined samples of various tumours in adults, 3 of which were samples of the STS group similar to the ones developing in children (angiosarcoma, liposarcoma, malignant fibrous histiocytoma). All tumours of a vascular background (haemangioma, lymphangioma, angiosarcoma) were positive for S1P₁. However, sensitivity and specificity in angiosarcoma tissues were much higher in comparison to a commonly used marker – CD31. This means that the evaluation of the S1P₁ receptor may be crucial for the correct differentiation of angiosarcoma with other tumours [36].

A different aspect of bioactive lipids' influence on neoplastic cells was researched by *Nofiele et al.* They revealed that in many of these cells – including mouse fibrosarcoma – ultrasound-stimulated microbubbles cause the increase of ceramides, which make these cells more sensitive to radiotherapy. This effect was nullified by exposing the abovementioned cells to physiological concentrations of S1P. Based on previous data, it can be concluded that the anti-apoptotic effect of S1P is directly linked with cell resistance to radiotherapy [37].

CONCLUSIONS

The role of bioactive lipids, including S1P and C1P, in the biology of neoplastic cells is unquestionable. All the

above-mentioned publications show various possibilities for improving the diagnostics and therapy of STS, and enable a better understanding of the mechanism of bioactive lipids' activity in both healthy and neoplastic cells. It is without doubt an aspect offering new therapeutic options for children with STS.

REFERENCES

- Miller R.W., Young J.L., Novacowic B.: Childhood cancer. *Cancer*. 1995, 75 (1 suppl.), 395–405.
- Paulino A.C., Okcu M.F.: Rhabdomyosarcoma. *Curr Probl Cancer*. 2008, 32 (1), 7–34.
- De Giovanni C., Landuzzi L., Nicoletti G., Lollini P.L., Nanni P.: Molecular and cellular biology of rhabdomyosarcoma. *Future Oncol*. 2009, 5 (9), 1449–1475.
- Toro J.R., Travis L.B., Wu H.J., Zhu K., Fletcher C.D., DeVesa S.S.: Incidence patterns of soft tissue sarcomas, regardless of primary site, in the surveillance, epidemiology and end results program, 1978–2001: An analysis of 26,758 cases. *Int J Cancer*. 2006, 119 (12), 2922–2930.
- Hannun Y.A., Obeid L.M.: Principles of bioactive lipid signaling: lessons from sphingolipids. *Nat Rev Mol Cell Biol*. 2008, 9 (2), 139–150.
- Hla T.: Physiological and pathological actions of sphingosine-1-phosphate. *Semin Cell Dev Biol*. 2004, 15 (5), 513–520.
- Chalfant C.E., Spiegel S.: Sphingosine-1-phosphate and ceramide 1-phosphate: expanding roles in cell signaling. *J Cell Sci*. 2005, 118, 4605–4612.
- Hinkovska-Galcheva V., Boxer L.A., Kindzelski A., Hiraoka M., Abe A., Goparaju S. et al.: Ceramide-1-phosphate a mediator of phagocytosis. *J Biol Chem*. 2005, 280 (28), 26612–26621.
- Germinario E., Peron S., Toniolo N., Betto R., Cencetti F., Donati C. et al.: S1P₂ receptor promotes mouse skeletal muscle regeneration. *J Appl Physiol*. 2012, 113 (5), 707–713.
- Gangoiti P., Bernacchioni C., Donati C., Cencetti F., Ouro A., Gomez-Munoz A. et al.: Ceramide-1-phosphate stimulates proliferation of C2C12 myoblasts. *Biochimie*. 2012, 94 (3), 597–607.
- Hokin M.R., Hokin L.E.: Enzyme secretion and the incorporation of P32 into phospholipids of pancreas slices. *J Biol Chem*. 1953, 203 (2), 967–977.
- Kihara A., Mitsutake S., Mizutani Y., Igarashi Y.: Metabolism and biological functions of two phosphorylated sphingolipids, sphingosine 1-phosphate and ceramide 1-phosphate. *Prog Lipid Res*. 2007, 46 (2), 126–144.
- Simons K., Toomre D.: Lipid rafts and signal transduction. *Nat Rev Mol Cell Biol*. 2000, 1 (1), 31–39.
- Ramstedt B., Slotte J.P.: Sphingolipids and the formation of sterol-enriched ordered membrane domains. *Biochim Biophys Acta*. 2006, 1758 (12), 1945–1956.
- Linn S.C., Kim H.S., Keane E.M., Andras L.M., Wang E., Merrill A.H.: Regulation of de novo sphingolipid biosynthesis and the toxic consequences of its disruption. *Biochem Soc Trans*. 2001, 29 (6), 831–835.
- Okazaki T., Bell R., Hannun Y.: Sphingomyelin turnover induced by vitamin D₃ in HL-60 cells. Role in cell differentiation. *J Biol Chem*. 1989, 264 (32), 19076–19080.
- Taha T.A., Argraves K.M., Obeid L.M.: Sphingosine-1-phosphate receptors: receptor specificity versus functional redundancy. *Biochim Biophys Acta*. 2004, 1682 (1–3), 48–55.
- Graler M.H., Bernhardt G., Lipp M.: S1P₄ receptor mediates S1P-induced vasoconstriction in normotensive and hypertensive rat lungs. *Pulm Circ*. 2011, 1 (3), 339–404.
- Kluk M.J., Hla T.: Signaling of sphingosine-1-phosphate receptors: receptor specificity versus functional redundancy. *Biochim Biophys Acta*. 2002 (1–3), 1582, 72–80.
- Brinkmann V.Y.: FTY 720: mechanism of action and potential benefit in organ transplantation. *Med J*. 2004, 45 (6), 991–997.
- Kim C.H., Wan W., Rui L., Kucia M., Laughlin M.J.: A novel paradigm in stem cell trafficking: the ratio of peripheral blood sphingosine-1-phosphate (S1P) to bone marrow ceramide-1-phosphate (C1P) regulates mobilization and homing of hematopoietic stem cells. *ASH meeting*. 2010 Abstract #554.

22. Lee M.J., Thangada S., Claffey K.P., Ancellin N., Liu C.H., Kluk M. et al.: Vascular endothelial cell adherens junction assembly and morphogenesis induced by sphingosine-1-phosphate. *Cell.* 1999, 99 (3), 301–312.
23. Spiegel S., Milstien S.: Sphingosine-1-phosphate: an enigmatic signaling lipid. *Nat Rev Mol Cell Biol.* 2003, 4 (5), 397–407.
24. Boudker O., Futerman A.H.: Detection and characterization of ceramide-1-phosphate activity in rat liver plasma membrane. *J Biol Chem.* 1993, 268 (29), 22150–22155.
25. Pappo A.S., Shapiro D.N., Crist W.M., Maurer H.M.: Biology and therapy of pediatric rhabdomyosarcoma. *J Clin Oncol.* 1995, 13 (8), 2123–2139.
26. Ferrari A., Bisogno G., Macaluso A., Casanova M., D'Angelo P., Pierani P. et al.: Soft-tissue sarcomas in children and adolescents with neurofibromatosis type 1. *Cancer.* 2007, 109 (7), 1406–1412.
27. Ruymann F.B., Maddux H.R., Ragab A., Soule L.H., Palmer N., Beltangady M. et al.: Congenital anomalies associated with rhabdomyosarcoma: an autopsy study of 115 cases. A report from The Intergroup Rhabdomyosarcoma Study Committee (representing The Children's Cancer Study Group, The Pediatric Oncology Group, The United Kingdom Children's Cancer Study Group, and The Pediatric Intergroup Statistical Center). *Med Pediatr Oncol.* 1988, 16 (1), 33–39.
28. Hennekam R.C.: Costello syndrome: an overview. *Am J Med Genet.* 2003, 117 (1), 42–48.
29. Tsokos M., Webber B.L., Parham D.M., Wesley R.A., Miser A., Miser J.S. et al.: Rhabdomyosarcoma. A new classification scheme related to prognosis. *Arch Pathol Lab Med.* 1992, 116 (8), 847–855.
30. Weiss A.R., Lyden E.R., Anderson J.R., Hawkins D.S., Spunt S.L., Walterhouse D.O. et al.: Histologic and clinical characteristic can guide staging evaluations for children and adolescents with rhabdomyosarcoma: a report from The Children's Oncology Group Soft Tissue Sarcoma Committee. *J Clin Oncol.* 2013, 31 (26), 3226–3232.
31. Bisogno G., Compostella A., Ferrari A., Pastore G., Cecchetto G., Garaventa A. et al.: Rhabdomyosarcoma in adolescents: a report from The AIEOP Soft Tissue Sarcoma Committee. *Cancer.* 2012, 118 (3), 821–827.
32. Crist W., Gehan E.A., Ragab A.H., Dickman D.S., Donaldson S.S., Fryer C. et al.: The Third Intergroup Rhabdomyosarcoma Study. *J Clin Oncol.* 1995, 13 (3), 610–630.
33. Raney R.B., Walterhouse D.O., Meza J.L., Andrassy R.J., Breneman J.C., Crist W.M. et al.: Results of The Intergroup Rhabdomyosarcoma Study Group D9602 protocol, using vincristine and dactinomycin with or without cyclophosphamide and radiation therapy, for newly diagnosed patients with low-risk embryonal rhabdomyosarcoma: a report from The Soft Tissue Sarcoma Committee of the Children's Oncology Group. *J Clin Oncol.* 2011, 29 (10), 1312–1318.
34. Schneider G., Bryndza E., Abdel-Latif A., Ratajczak J., Maj M., Tarnowski M. et al.: Bioactive lipids sphingosine-1-phosphate and ceramide-1-phosphate are pro-metastatic factors in human rhabdomyosarcomas cell lines, and their tissue level increases in response to radio/chemotherapy. *Mol Cancers Res.* 2013, 11 (7), 793–807.
35. Schneider G., Sellers Z.P., Abdel-Latif A., Morris A.J., Ratajczak M.Z.: Bioactive lipids, LCP and LPA are novel prometastatic factors and their tissue levels increase in response to radio/chemotherapy. *Mol Cancer Res Published Online First July 2014; DOI: 10.1158/1541-7786. MCR-14-0188.*
36. Akiyama T., Hamazaki S., Monobe Y., Nishimura H., Irei I., Sadahira Y.: Sphingosine 1-phosphate receptor 1 is a useful adjunct for distinguishing vascular neoplasms from morphological mimics. *Virchows Arch.* 2009, 454 (2), 217–222.
37. Nofiele J.T., Karshafian R., Furukawa M., Al-Mahrouki A., Giles A., Wong S. et al.: Ultrasound-activated microbubble cancer therapy: ceramide production leading to enhanced radiation effect in vitro. *Technol Cancer Res Treat.* 2013, 12 (1), 53–60.